



THE UNIVERSITY OF  
**AUCKLAND**  
Te Whare Wānanga o Tāmaki Makaurau  
NEW ZEALAND

# MicroRNAs

## Sequencing, analysis ... and then what?

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Nicole Cloonan - 4<sup>th</sup> July 2017

#UQWinterSchool #Bioinformatics #GroupTherapy

Woohoo! I'm going to do a miRNAseq project!



Here are some things to think about.



OMFG – my career is over.



# The five stages of miRNA analysis

Denial

OMG – I have started an amazing project, this will get me a Nobel Prize for sure!

Ancient Greek word

“little”

Ancient Greek word

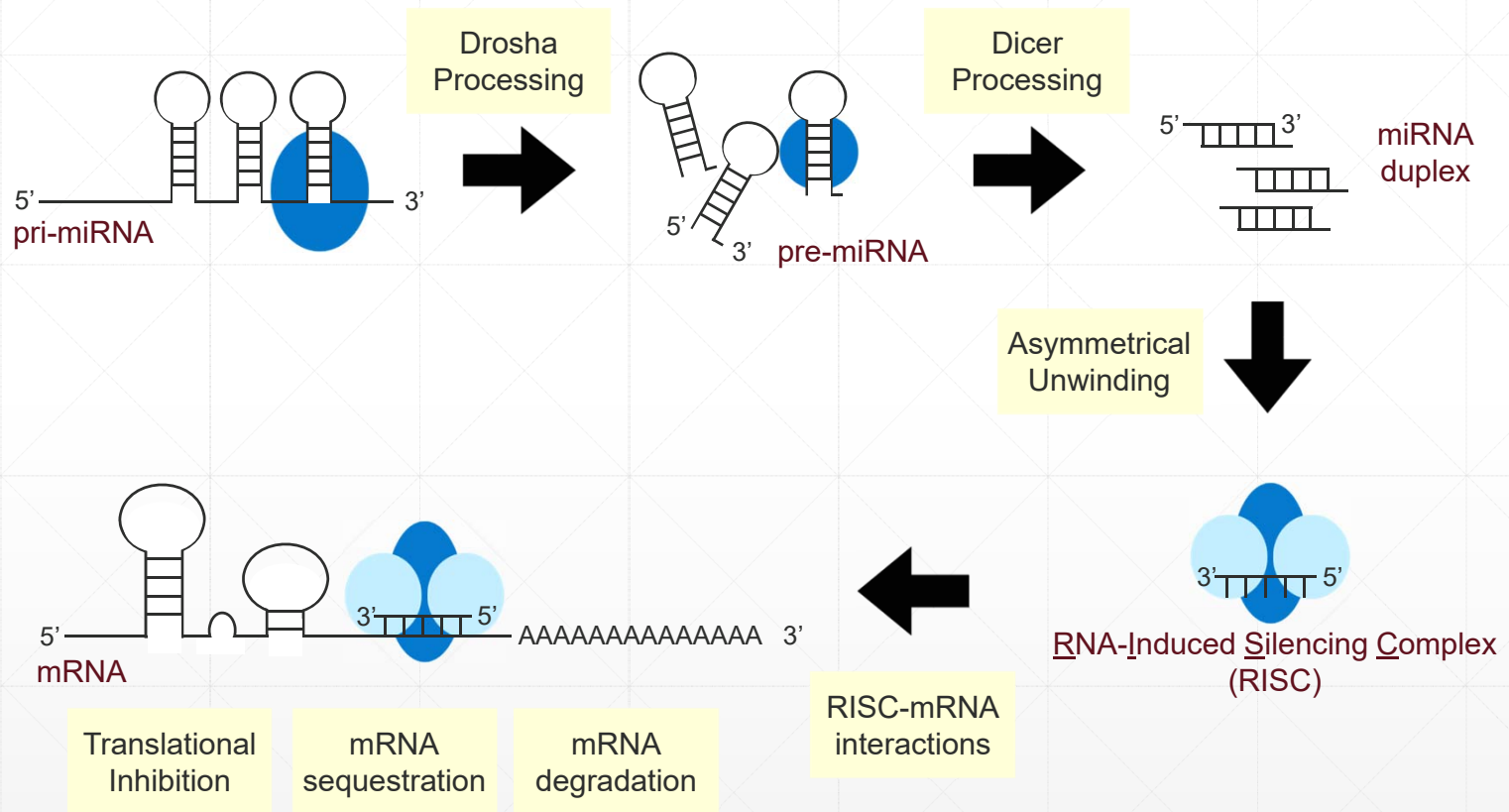
“bastard”

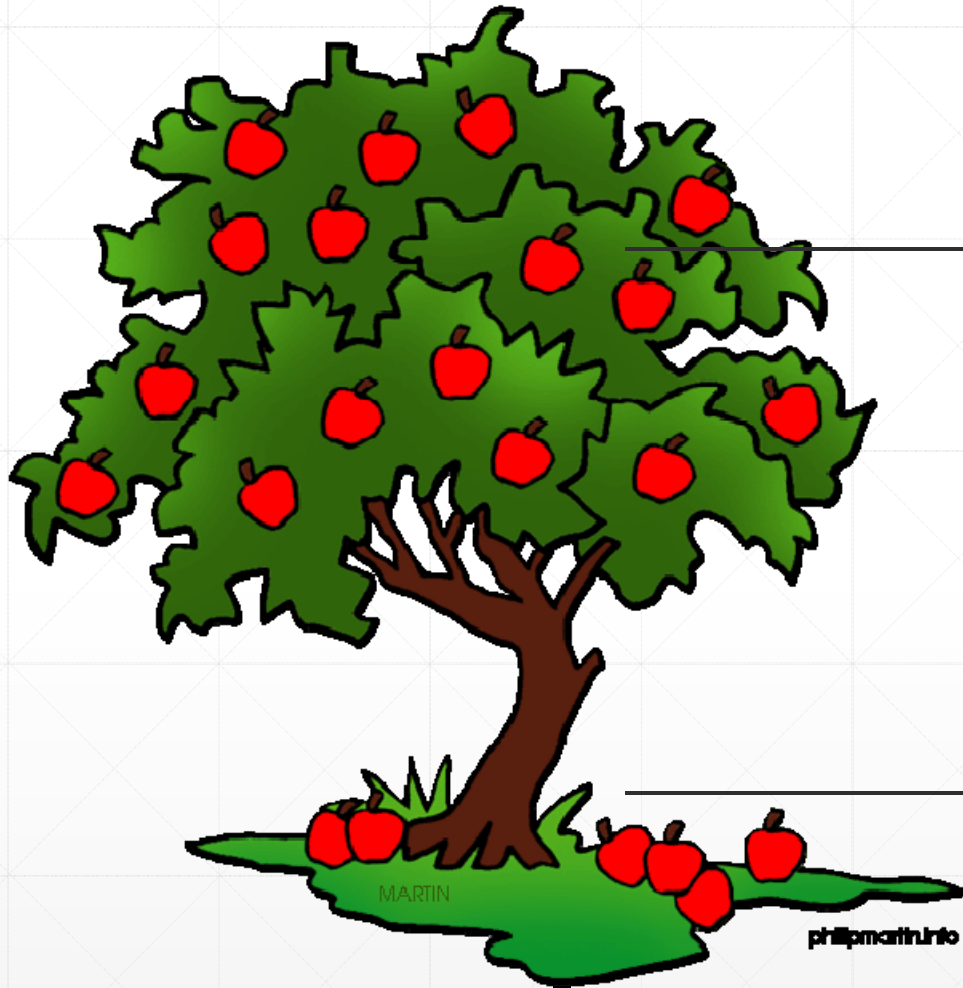
# microRNA

noun mi·cro·RNA \,mī-krō-,är-(,)en-'ā\

variants: **miRNA** or less commonly **micro-RNA**

# miRNA biogenesis





## Low Hanging Fruit

- Cheap and easy to sequence (~\$80 per sample)
- A natural fit for short read sequencing technologies
- Fantastic possibilities as biomarkers (stable)
- Interesting possibilities as therapeutics
- Established modes of action
- Enables lazy department store science

## Ground Fruit

- Flawed, unpalatable, and a probably a little smelly

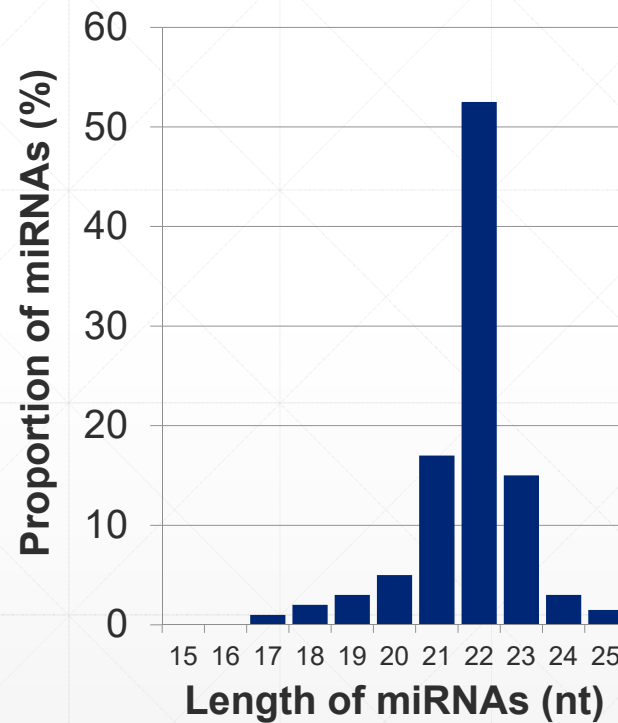
# The five stages of miRNA analysis

Anger

What do you  
mean we can't  
align these  
things well?

# Anger – the project is not as easy as your supervisor made you think it was

miR-17-5p : CAAAGUGCUUACAGUGCAGGUAGU  
miR-20 : UAAAGUGCUUAUAGUGCAGGUAG-  
miR-106a : AAAAGUGCUUACAGUGCAGGUAGC  
miR-106b : UAAAGUGCUGACAGUGCAGAU---  
miR-93 : -AAAGUGCUGUUCGUGCAGGUAG-  
miR-18 : UAAGGUGCAUCUAGUGCAGAUA--  
  
miR-19a : UGUGCAAUCCUAUGCAAACUGA-  
miR-19b-1 : UGUGCAAUCCUAUGCAAACUGA-  
miR-19b-2 : UGUGCAAUCCUAUGCAAACUGA-



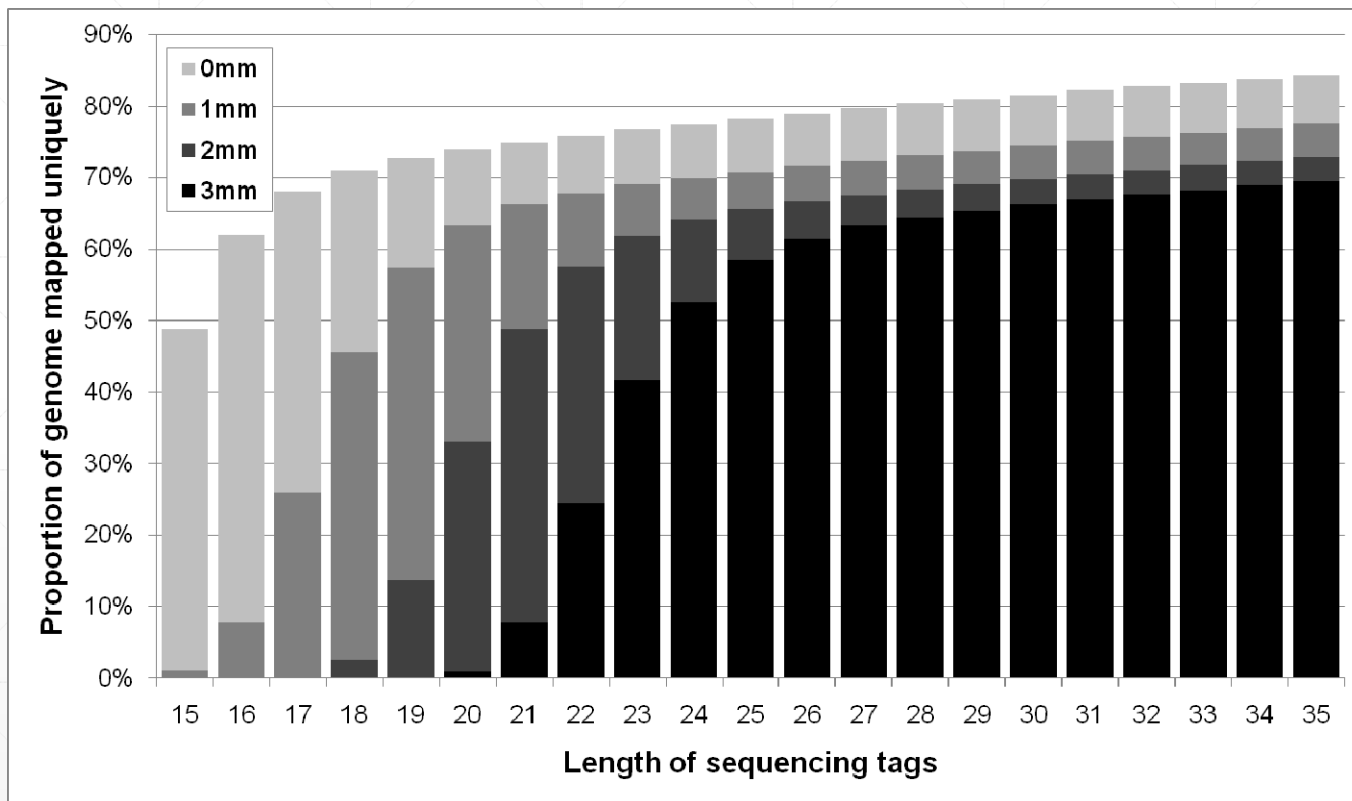


## The two laws of repeats

1. It is impossible to resolve repeats of length  $L$  unless you have reads longer than  $L$ .
2. It is impossible to resolve repeats of length  $L$  unless you have reads longer than  $L$ .

Gleefully stolen from  
[@torstenseemann](#)

# Aligning to the genome



## Allowing errors in alignments

- Your reference genome will contain polymorphisms
- Your sample will contain amplification or sequencing artefacts
- Your sample may contain RNA editing
- Unique alignment is not the same as specific alignment
- Therefore, you must allow at least as many errors per alignment as you are expecting in your data

## Watch out for your aligner!

- Make sure your aligner and/or alignment parameters allow proper mapping of small RNAs.

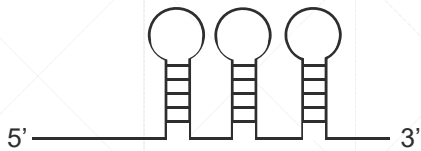
miRNA size = 22nt

caaagugcuuacagugcaggua

Seed size = 16nt



# Alternatives to genome alignment



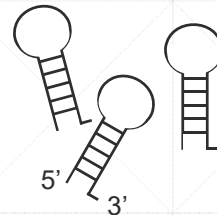
## pri-miRNA transcript

Could also be pre-mRNA transcript with miRNAs in the introns

1000s nt long

Often not fully defined

Will miss novel miRNAs



## pre-miRNA hairpins

80-200 nt long

Will miss novel miRNAs



## Mature miRNAs

18-25 nt long

Will miss novel miRNAs

Will not detect isomiRs

Best curated resource for miRNAs is currently miRbase (<http://miRBase.org>)

# The five stages of miRNA analysis

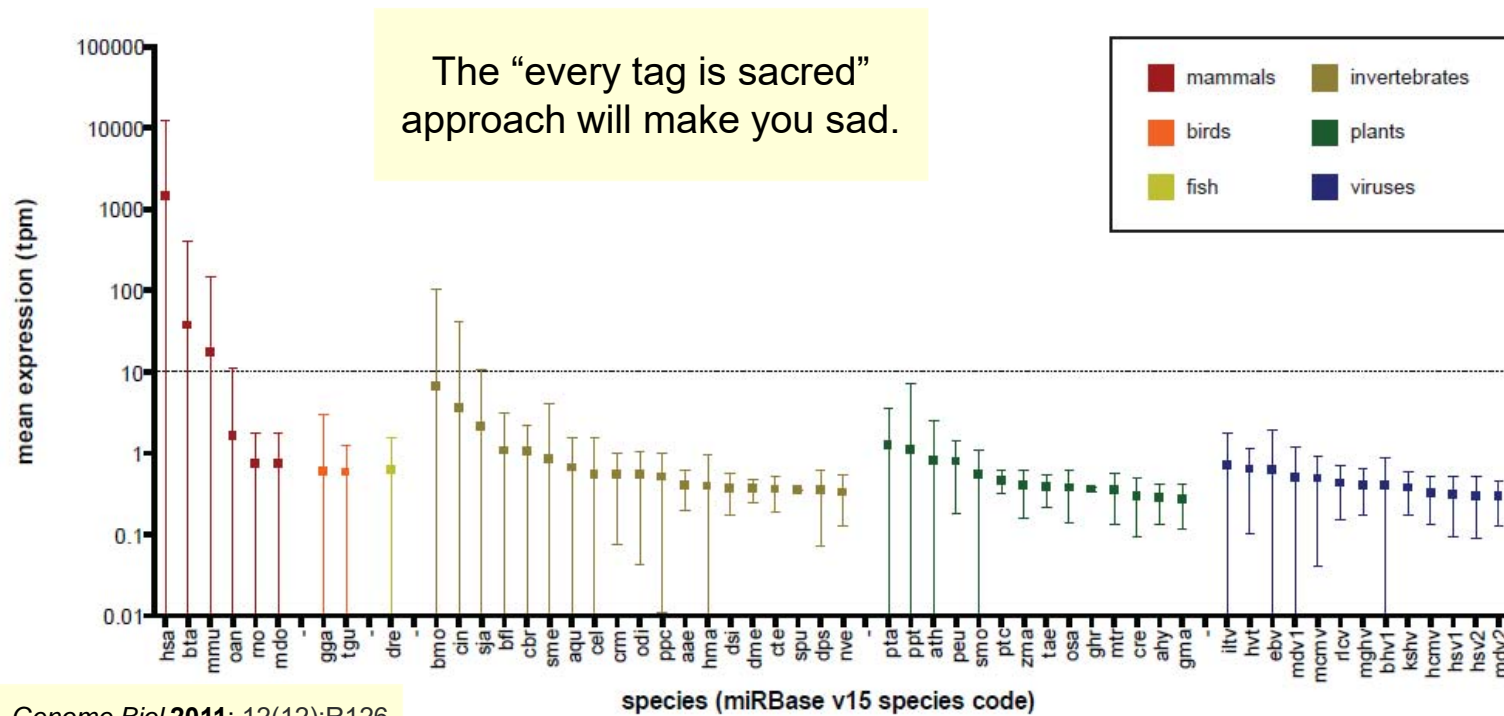
**Bargaining**

Can we please  
have some  
more  
replicates?

## Bargaining – maybe I can still get useful data if I do the analysis well

1. What miRNAs are expressed in my sample?
2. What miRNAs are different between my samples?

# What miRNAs are expressed?



Cloonan *et al. Genome Biol* 2011; 12(12):R126



# What miRNAs are different?

- You absolutely **CANNOT** answer this question without replicates
  - All biological systems are variable – it's an evolutionary **NEED**
  - Understanding biological variations means no wasted time chasing false positives
  - Your question is actually whether the variation between your groups is bigger than the variation within your groups
- You must have independent biological replicates
  - Do not fall into the “reducing variation” trap
  - Variation is your friend, not your enemy – you want to **MAXIMIZE** your biological variation
  - Using technical replicates makes your p-values **WORTHLESS**
- If you can't afford replicates – then you can't afford the experiment.

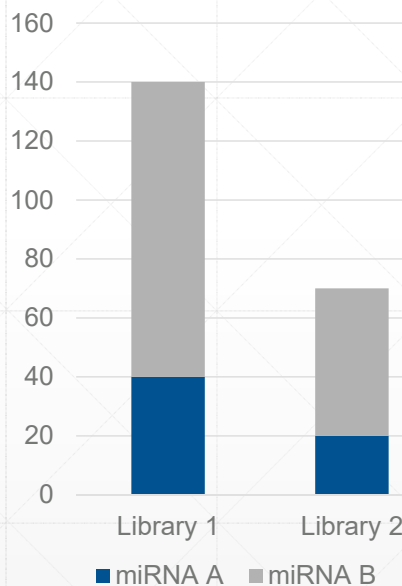
# Comparing groups - Normalization

- “Housekeeping Genes”
  - Could we all please agree that this is not a thing and to stop bringing it up?
  - You shouldn’t even be doing this for qRT-PCR let alone sequencing
- Spike-Ins
  - One potential method of normalization, but be careful
  - There is error when measuring any individual transcript, when you “correct” with a spike-in, you multiply the spike-in error with the error of each transcript
  - A spike in added as a percent of the RNA instead of a percent of the cells will mask any global changes in RNA production or composition.
- Data-driven normalization (eg. Quantile normalization)
  - Be careful – relies on the assumption that the vast majority of genes/miRNAs will not change
  - Only a few thousand data points for these approaches to model – check software assumptions

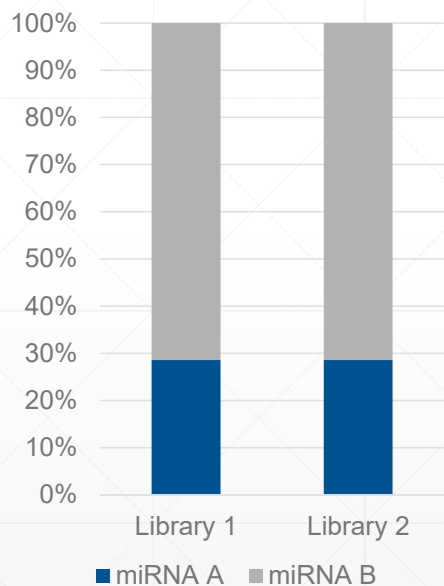
# Scaling to Library Depth

TMM – Trimmed Mean of M Values

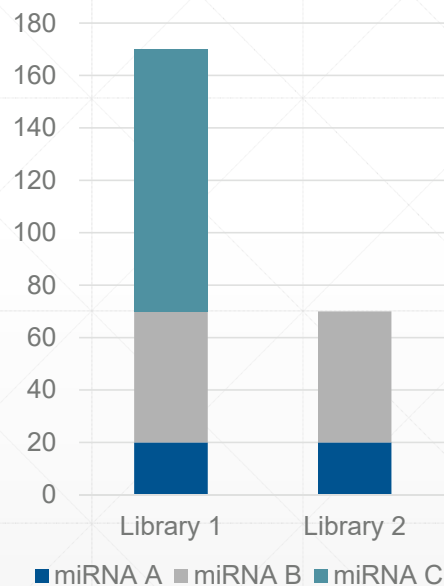
Raw Read Counts



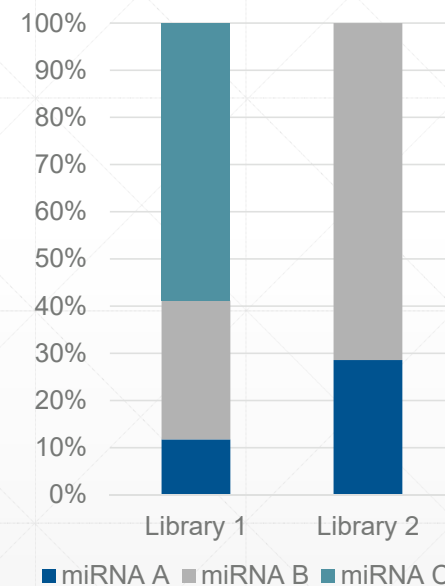
Counts per Million



Raw Read Counts

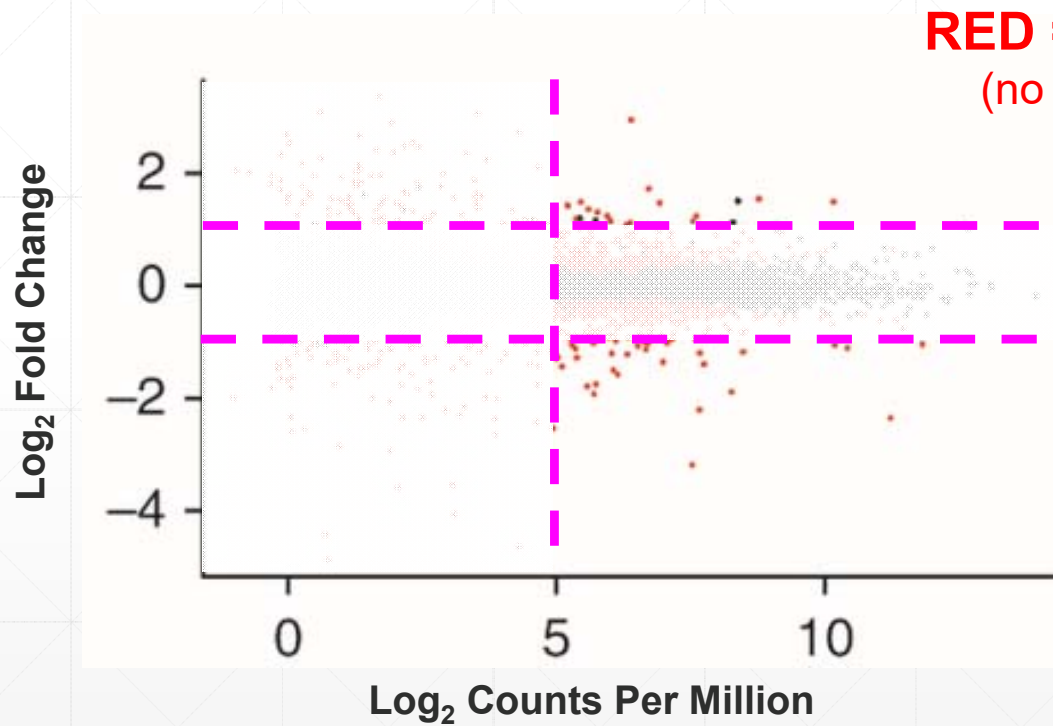


Counts per Million



Robinson & Oshlack *Genome Biol* 2010; 11(3):R25

# What miRNAs are different?



**RED = statistical significance**  
(no replicates = no p-values)

# The five stages of miRNA analysis

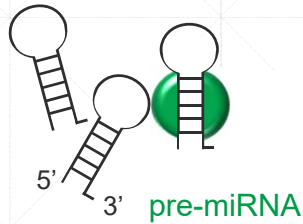
Depression

OMFG – do  
these miRNA  
problems never  
end?

## Depression - you have a list of miRNAs but realise that no one cares

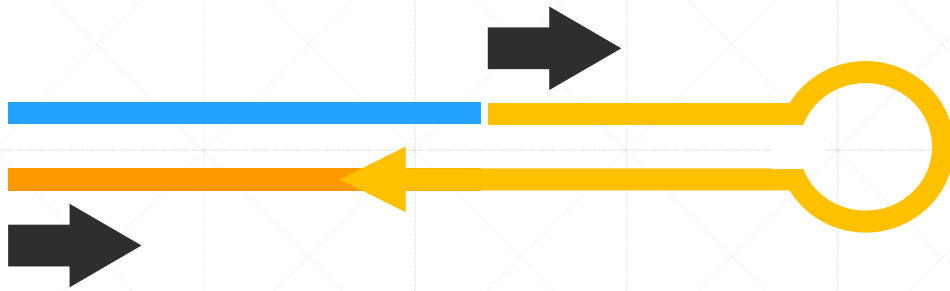
- If you are doing a biomarker study, then you need to demonstrate utility
- If you are doing a mechanistic study, then you need to demonstrate function

# IsomiRs and biomarker validation

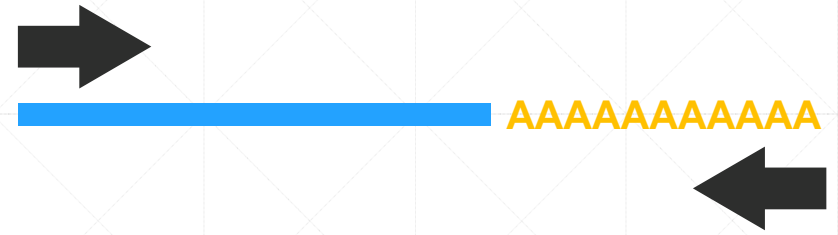


Cloonan *et al. Genome Biol* **2011**; 12(12):R126

# qRT-PCR and IsomiRs



This method is isomiR specific



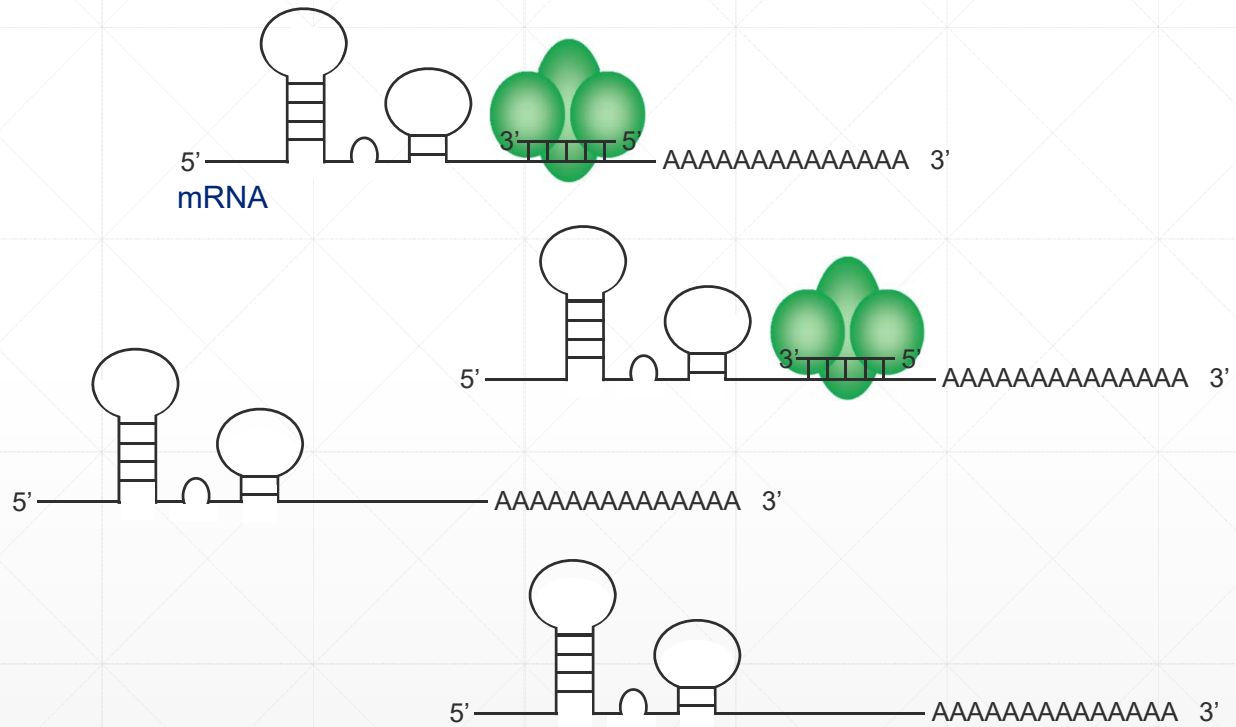
This method is isomiR agnostic



## Mechanism - What is a target anyway?

- miRNAs can bind to mRNAs without inducing a repressive effect
  - This happens with transcription factors due to inherent non-functioning sites, or limiting co-factor concentrations
- Full or partial repression of a protein/mRNA does not necessarily alter the trajectory of a cell
  - Different molecules have different tolerances of variability in expression
- Full or partial repression does not mean a change in protein/mRNA levels
  - Long half lives, stoichiometry

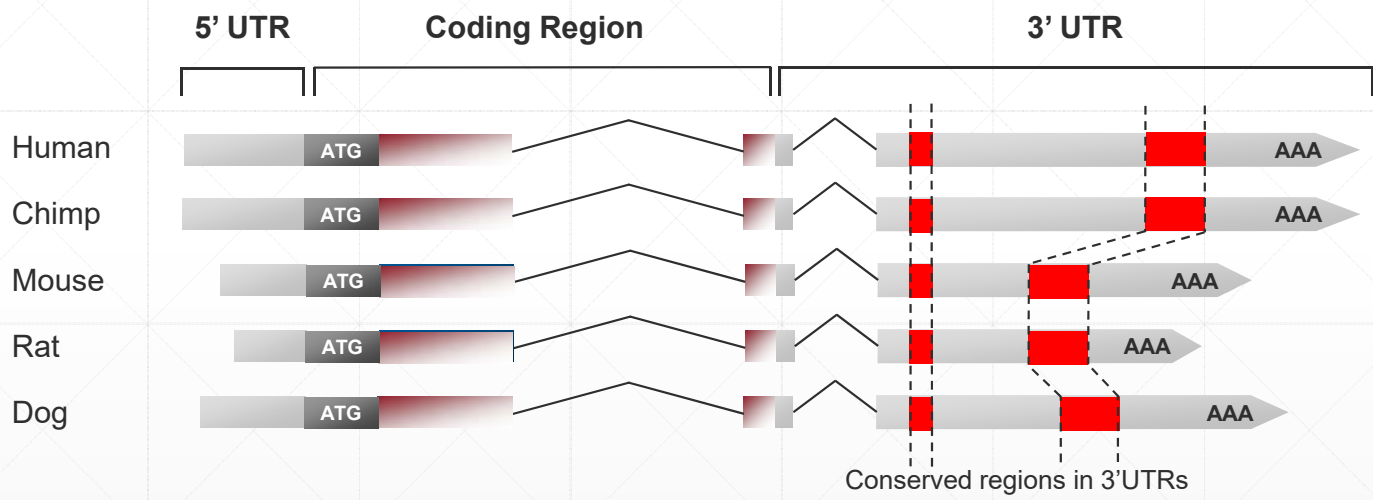
# Stoichiometry of miRNA Interactions



Cloonan *et al.*, *Genome Biology*, 2008 9:R127

# Bioinformatics methods for target PREDICTION

Assumptions have to be made to achieve any kind of specificity

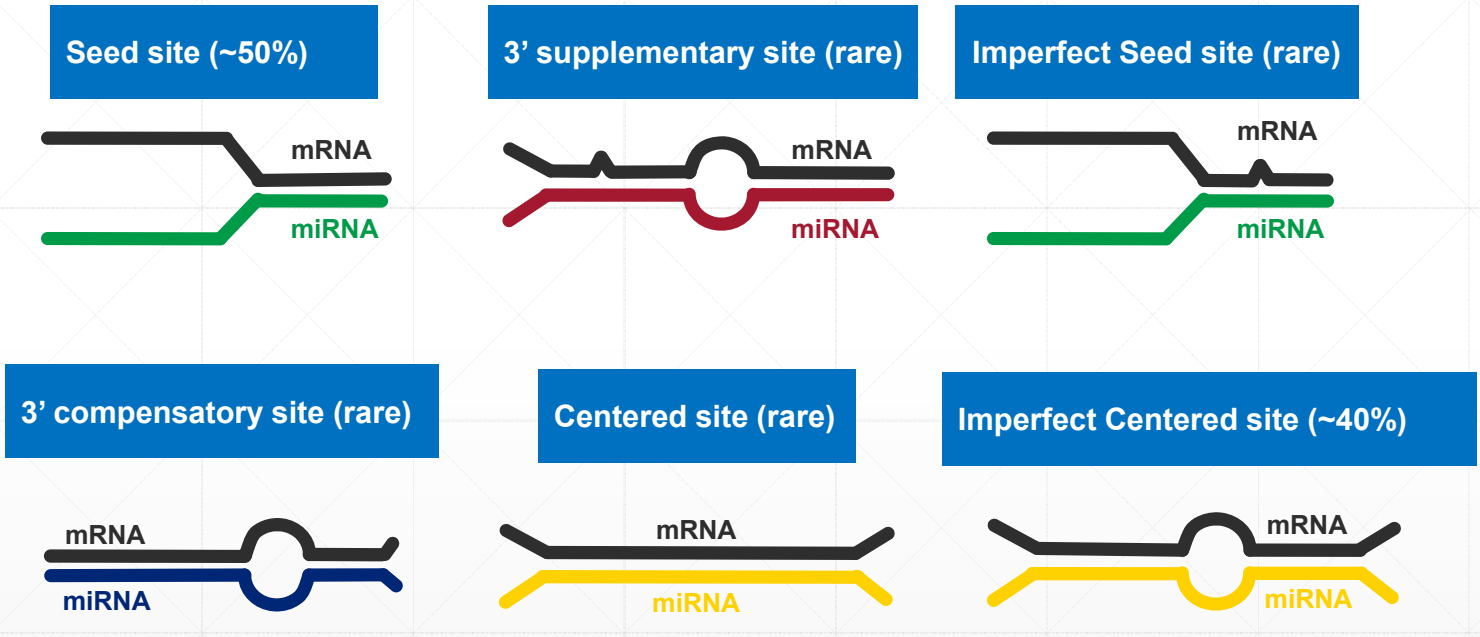


Best have 50% false positive rates, and at least 40% false negative rates

## The dominance of the seed site

- Nucleotides 2-8 in the 5' end of the miRNA are called the “seed” site.
  - Fun fact: first miRNA interactions were not seed sites
  - Fun fact: seed sites have the most predictive value. This is NOT the same thing as being the most functional
- Literature bias towards seed sites because experimental validation focuses around those most likely to give the “best” results for the lowest time/cost investment

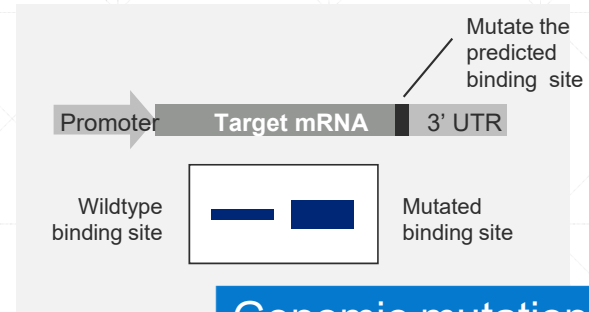
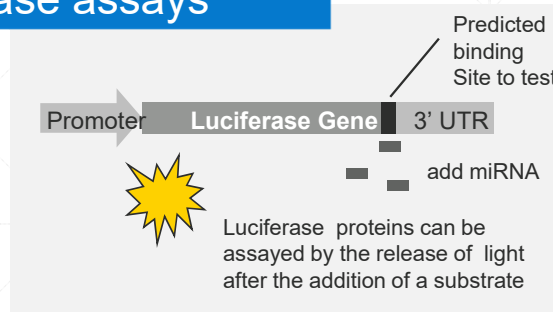
# Modes of miRNA binding



Adapted from Brennecke *et al.*, (2005) PLoS Biology 3:e85

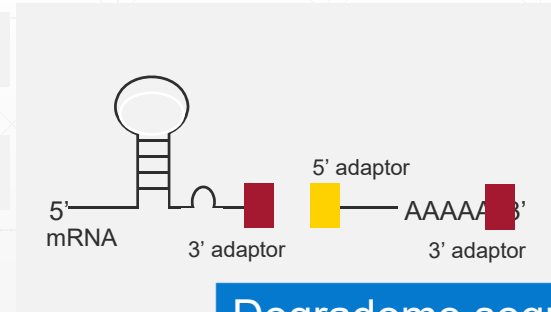
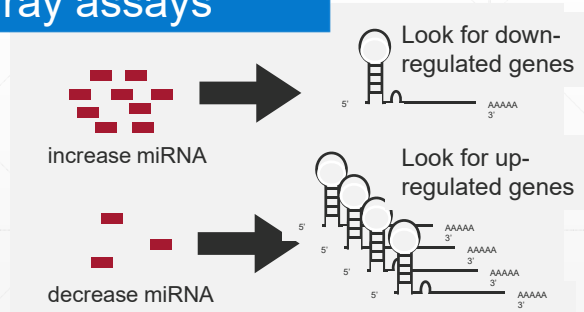
# Target detection methods

## Luciferase assays



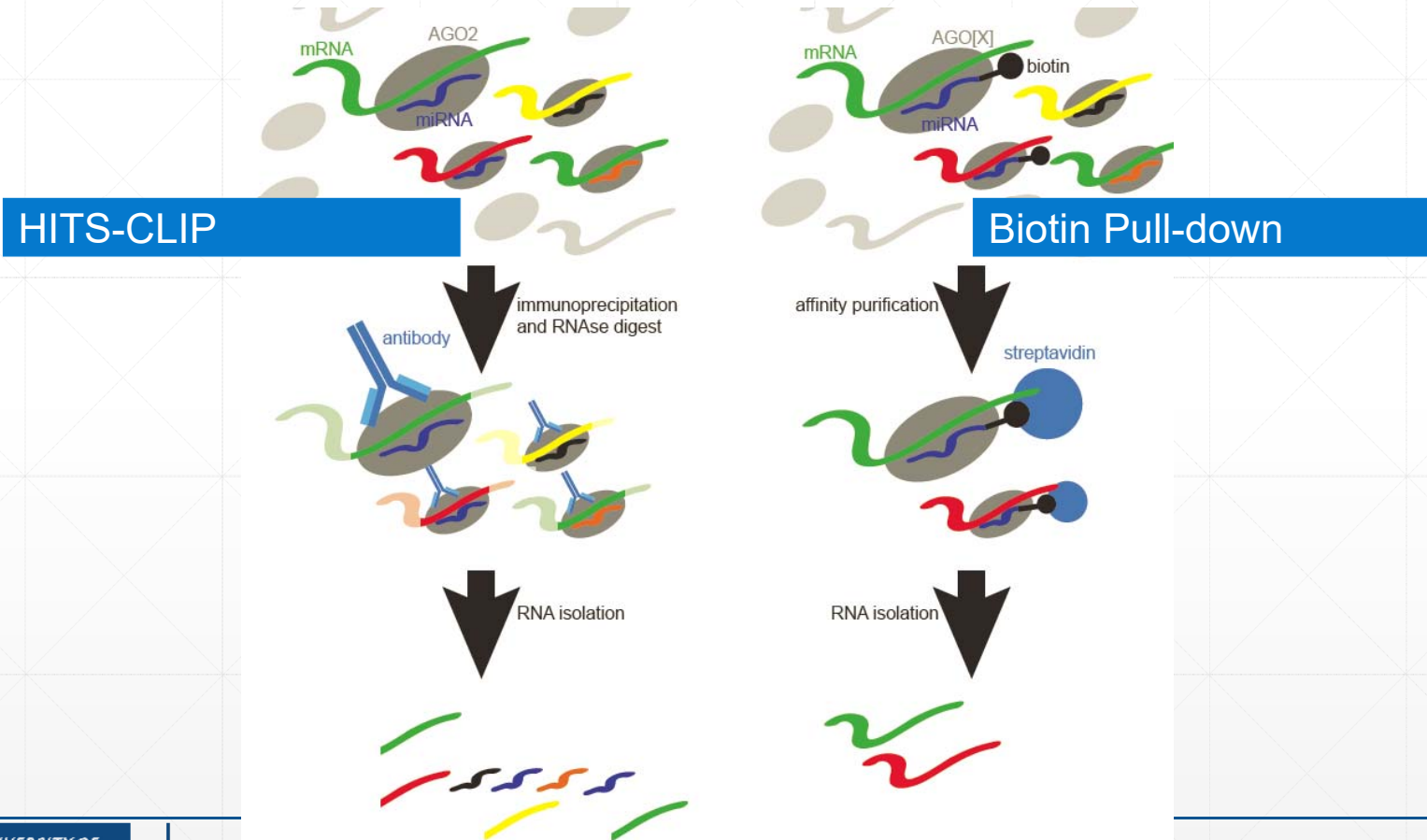
## Genomic mutation

## Microarray assays



## Degradome sequencing

# Target detection methods



# The five stages of miRNA analysis

Acceptance

OK, it's not as amazing as I thought, but it's probably still possible to write a decent paper.



## Conclusions

- Seriously – use replicates. No, really. Do it.
- Check your assumptions. No, really. Do it.
- Methods matter, understand what each method tells you... and what it DOESN'T

**MicroRNAs can be a fun project, but are not an easy project. Exercise caution and don't over interpret your results.**

## Last Word...

99.5% of people with a biology PhD will not retire while still in a research role. Hard work and talent are not sufficient to guarantee a long career. Your time in research may be short, but your contribution to the literature is FOREVER.

So take the time to do GOOD science.

# Our Time is Up!



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# A parable of miRNA function

